

Association of 41 circulating inflammatory factors, C-reactive protein, and procalcitonin with sepsis risk and 28-day mortality: A bidirectional Mendelian randomization and mediation analysis

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Advances in Clinical and Experimental Medicine, ISSN 1899–5276 (print), ISSN 2451–2680 (online)

Adv Clin Exp Med. 2026;35(1):45–55

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Funding sources

This work was supported by the Guangxi Science and Technology Department Project (grant No. Gui Ke AB23026098), the Guangxi Science and Technology Department Project (grant No. GZKJ2307), and the National Natural Science Foundation of China (grant No. 82305151).

Conflict of interest

None declared

Acknowledgements

The authors express their sincere gratitude to the researchers who generously shared the GWAS data used in this study.

Received on July 14, 2024

Reviewed on February 8, 2025

Accepted on March 20, 2025

Published online on January 5, 2026

Abstract

Background. The precise causal relationship between circulating inflammatory factors and sepsis has not yet been fully elucidated.

Objectives. To identify biomarkers that enable earlier and more accurate diagnosis of sepsis.

Materials and methods. The causal relationships between 41 circulating inflammatory factors, C-reactive protein (CRP), and procalcitonin (PCT) with sepsis and 28-day sepsis-related mortality were evaluated using two-sample bidirectional Mendelian randomization (MR) analyses.

Results. This study revealed negative causal associations between genetically predicted circulating inflammatory factors—interleukin-6 (IL-6) (odds ratio [OR] = 0.923; 95% confidence interval [CI], 0.854–0.998; $p = 0.044$), RANTES (OR = 0.926; 95% CI, 0.862–0.994; $p = 0.033$), and macrophage inflammatory protein-1 β (MIP1 β) (OR = 0.957; 95% CI, 0.919–0.996; $p = 0.032$)—and the risk of sepsis. Furthermore, positive causal relationships were observed between CRP and sepsis (OR = 1.140; 95% CI: 1.055–1.232; $p = 0.001$), as well as between CRP and 28-day sepsis-related mortality (OR = 1.241; 95% CI: 1.034–1.489; $p = 0.020$). Platelet-derived growth factor-BB (PDGF-BB) levels were also elevated in sepsis (OR = 1.136; 95% CI: 1.003–1.286; $p = 0.044$). Mediation analysis indicated that CRP mediated the effects of IL-6 and RANTES on sepsis, accounting for 25.87% (OR = 0.980; 95% CI: 0.961–0.998) and 2.04% (OR = 1.002; 95% CI: 0.991–1.012) of the total effect, respectively. The robustness of these associations was confirmed through leave-one-out sensitivity analysis and funnel plots.

Conclusions. This study enhances our understanding of the mechanisms underlying sepsis and its associated mortality, and underscores the therapeutic potential of targeting inflammatory factors in its management.

Key words: C-reactive protein, procalcitonin, sepsis, Mendelian randomization analysis, circulating inflammatory factors

Cite as

Peng J, Li L, Wang G, et al. Association of 41 circulating inflammatory factors, C-reactive protein, and procalcitonin with sepsis risk and 28-day mortality: A bidirectional Mendelian randomization and mediation analysis.

Adv Clin Exp Med. 2026;35(1):45–55.

doi:10.17219/acem/203155

DOI

10.17219/acem/203155

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Highlights

- The study investigated causal relationships between 41 circulating inflammatory factors, including C-reactive protein (CRP) and procalcitonin (PCT) – and the risk of sepsis and 28-day sepsis-related mortality using Mendelian randomization analysis.
- Genetically predicted inflammatory mediators, including interleukin-6 (IL-6), RANTES, and macrophage inflammatory protein-1 β (MIP-1 β), showed negative causal associations with sepsis risk.
- Positive causal relationships were identified between CRP and both sepsis incidence and 28-day sepsis-associated mortality.
- Platelet-derived growth factor-BB (PDGF-BB) levels were found to be significantly elevated in patients with sepsis.
- CRP acted as a mediating factor linking IL-6 and RANTES to the development of sepsis, suggesting a key pathway in sepsis pathophysiology.

Background

Sepsis is a syndrome characterized by a systemic inflammatory response triggered by infection of the host with pathogenic agents such as bacteria, viruses, fungi, or parasites, and it is often secondary to trauma, burns or shock.^{1–3} This condition frequently leads to dysfunction or failure of multiple organs, posing a serious threat to human life. It has been reported that the global burden of sepsis includes 48.9 million cases and 11 million sepsis-related deaths annually,⁴ while the overall incidence of sepsis among intensive care unit (ICU) patients in Asian countries is 22.4%.

The 30-day mortality rate of sepsis is approx. 17.4%, while that of severe sepsis can reach 26%, and may rise to 32.6% in low- and middle-income countries.^{5–7} Additionally, studies have reported that the incidence of sepsis can be as high as 38.6%, with mortality rates reaching 43.0% and continuing to increase at an alarming pace.^{8–10} Sepsis remains a major focus in the field of critical and emergency medicine, underscoring the urgent need to deepen our understanding of its pathophysiology, improve prevention strategies, and strengthen therapeutic approaches.

Identifying the etiology of sepsis is essential for reducing its incidence and improving treatment outcomes. The physiological and pathological processes of sepsis are highly complex, but the systemic inflammatory response remains one of its core pathological mechanisms. An excessive inflammatory reaction can lead to organ dysfunction in sepsis. Numerous preclinical studies have demonstrated significant increases in the protein levels of interleukin 6 (IL-6) and tumor necrosis factor alpha (TNF- α) in the liver tissue of septic rats.¹¹

Serum concentrations of TNF- α , IL-6 and IL-10 are markedly elevated in rats with sepsis.¹² In cellular models of sepsis, the secretion of IL-6, TNF- α and IL-1 β increases sharply, amplifying the inflammatory response.^{13,14} Clinical studies have confirmed these findings, demonstrating that procalcitonin (PCT) serves as a potential indicator of disease severity, whereas IL-6 is associated with

the clinical progression of the syndrome.¹⁵ Both PCT and C-reactive protein (CRP) are significantly upregulated in sepsis.¹⁶ Moreover, increased IL-27 levels and decreased RANTES concentrations have been observed in neonates with sepsis.¹⁷

Observational studies often carry a risk of bias and false associations. The Mendelian randomization (MR) approach addresses this limitation by using genetic variants as instrumental variables (IVs) to infer causality. Mendelian randomization reduces confounding factors and bias, providing stronger evidence for causal relationships.^{18,19} The two-sample MR method further minimizes distortion and interference by employing pooled statistical data from multiple studies. Moreover, this approach has been widely applied to improve causal inference through larger sample sizes and greater statistical power.

Objectives

To identify potential causal relationships and mediators between inflammatory factors and sepsis, this study employed a bidirectional MR design combined with mediation analysis. Additionally, the study aimed to provide guidance for the development of diagnostic and therapeutic strategies based on 41 circulating inflammatory factors (Table 1), as well as CRP and procalcitonin PCT, which may assist in the early diagnosis and treatment of sepsis.

Materials and methods

Research design

A genome-wide association study (GWAS) is a research approach used to identify genetic variants associated with specific diseases or traits. This method involves scanning the entire genome of a large number of individuals to detect genetic markers linked to the condition or

Table 1. Circulating inflammatory factors

Number	Inflammatory factors
1	interleukin 1 β
2	interleukin 1-receptor antagonist
3	interleukin 2
4	interleukin 2-receptor antagonist
5	interleukin 4
6	interleukin 5
7	interleukin 6
8	interleukin 7
9	interleukin 8
10	interleukin 9
11	interleukin 10
12	interleukin 12p70
13	interleukin 13
14	interleukin 16
15	interleukin 17
16	interleukin 18
17	cutaneous T-cell attracting
18	stromal-cell-derived factor 1 alpha
19	regulated on activation, normal T cell expressed and secreted
20	macrophage inflammatory protein 1 β
21	macrophage inflammatory protein 1a
22	monokine induced by interferon gamma
23	monocyte chemoattractant protein-3
24	monocyte chemoattractant protein-1
25	interferon γ -induced protein 10
26	growth-regulated protein a
27	eotaxin
28	beta-nerve growth factor
29	vascular endothelial growth factor
30	stem cell growth factor beta
31	stem cell factor
32	platelet-derived growth factor BB
33	macrophage colony stimulating factor
34	hepatocyte growth factor
35	granulocyte-colony stimulating factor
36	fibroblast growth factor basic
37	macrophage migration inhibitory factor
38	tumor necrosis factor-related apoptosis inducing ligand
39	tumor necrosis factor alpha
40	tumor necrosis factor beta
41	interferon gamma

characteristic of interest. The present study is based on the most recent and comprehensive GWAS datasets from large populations to investigate the causal relationships of CRP, PCT and 41 circulating inflammatory factors with sepsis and 28-day mortality.

To ensure the validity of the research findings, the IV model in MR must satisfy 3 key assumptions: 1) the genetic variants are strongly associated with the exposure variables; 2) the genetic variants are independent of confounding factors that may influence the results (e.g., smoking, diabetes); and 3) the genetic variants affect the outcome variables only through the exposure variables. The overall research process is illustrated in Fig. 1.

Collection of data on circulating inflammatory factors

Genetic instruments for the 41 circulating inflammatory factors were obtained from the most recent and comprehensive GWAS datasets, which combined summary statistics from 3 Finnish cohorts – the Cardiovascular Risk in Young Finns Study, FINRISK 1997, and FINRISK 2002 (n = 8,293);^{20,21} 13 European-ancestry cohorts (n = 21,758);²² and the Lothian Birth Cohort 1936 (n = 1,091).²³ The GWAS data on 41 circulating inflammatory factors involved a total of 31,142 individuals.

The GWAS data for CRP were obtained from the UK Biobank (n = 427,367) and the Cohorts for Heart and Aging Research in Genomic Epidemiology (CHARGE) consortium (n = 148,164), comprising a total of 575,531 participants.²⁴ The GWAS data for PCT were derived from 3,301 healthy participants from the INTERVAL study.²⁵

Further details regarding genomic (GM) statistics are available in the original publications. As indicated in Supplementary Table 1, the IEU OpenGWAS project also provides access to the corresponding GWAS datasets.

Collection of sepsis datasets

The OpenGWAS database includes summary statistics for sepsis and sepsis-related 28-day mortality, obtained from the IEU OpenGWAS project, which is based on data from the UK Biobank.²⁶ This dataset comprises 11,643 sepsis patients of European ancestry and 474,841 controls, with a total of 12,243,539 SNP loci analyzed. Patients were classified as having confirmed sepsis according to the International Classification of Diseases (ICD-9 and ICD-10) codes.⁴ Because data on circulating inflammatory factors and sepsis were obtained from different research projects, there was no sample overlap. As this study did not involve the collection of individual participant data, ethical approval was not required.

Selection of genetic instrumental variables

First, a strong correlation must exist between the exposure factors and the instrumental variables selected for analysis. To ensure adequate screening of instrumental variables, single-nucleotide polymorphisms (SNPs) with p-values below the genome-wide significance threshold (1×10^{-5}) were selected. Subsequently, the genetic distance

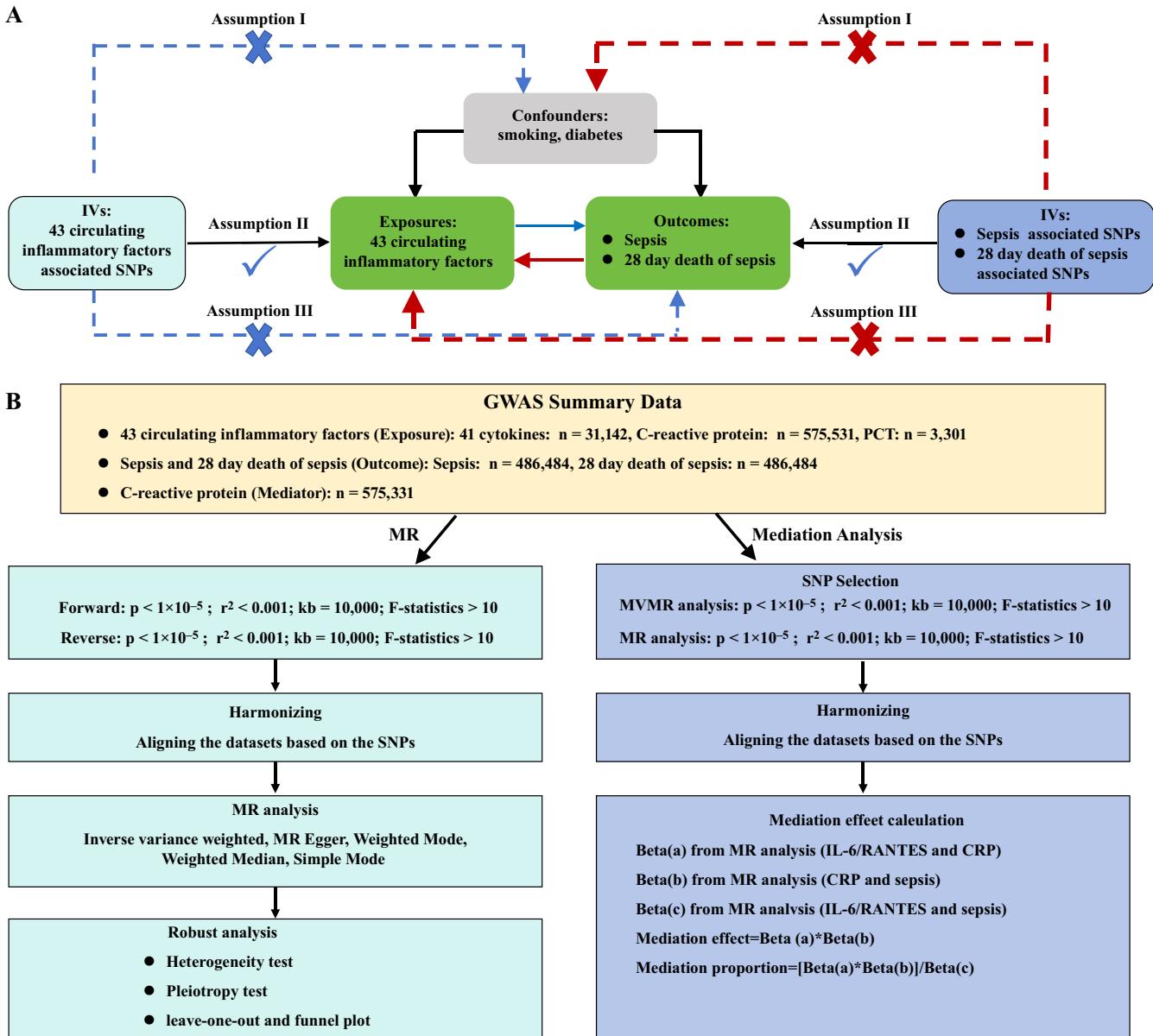


Fig. 1. The bidirectional Mendelian randomization (MR) and mediation analysis study of the associations between 41 circulating inflammatory factors, C-reactive protein (CRP), procalcitonin (PCT) and sepsis

was set to 10,000 kb, and the linkage disequilibrium (LD) parameter (R^2) for the SNPs was set to 0.001. Next, we calculated the F-statistic using the formula:

$$F = [(R^2/(1 - R^2)) \times (N - K - 1)/K]$$

Instrumental variables with F-values < 10 were excluded to ensure a strong correlation between the exposure factors and IVs. Additionally, IVs showing associations with outcome variables at a p -value $< 1.0 \times 10^{-5}$ were removed. The Phenoscanner database²⁷ (<http://www.phenoscanner.medschl.cam.ac.uk/>) was used to identify and exclude potential confounding factors (e.g., smoking, diabetes) related to the IVs. Finally, the exposure and outcome datasets were harmonized to remove non-concordant SNPs, leaving the remaining SNPs as valid genetic instrumental variables.

Bidirectional MR analysis

A bidirectional MR approach was used to assess the causal relationships between circulating inflammatory factors and sepsis. Mendelian randomization analysis was performed using the inverse-variance weighted (IVW) method as the primary analytical strategy. The IVW approach combines all SNPs in a meta-analysis framework to generate a pooled causal estimate. Four additional complementary methods – MR-Egger, weighted mode, weighted median, and simple mode – were also applied. Sensitivity analyses were conducted using the MR-Egger and weighted median methods. When IVs account for at least half of the total weight in the analysis, the weighted median method provides robust and reliable causal estimates.

Pleiotropy was assessed and corrected using the MR-Egger intercept test. Heterogeneity among SNPs selected for each exposure variable was evaluated by calculating Cochran's Q statistic. The Mendelian Randomization Pleiotropy RESidual Sum and Outlier (MR-PRESSO) method was applied to detect outliers and provide corrected causal estimates. The p-value from the MR-Egger intercept test was used to assess the presence of pleiotropy among SNPs. A nonsignificant difference between Q and Q' ($p > 0.05$) indicated that the IVW model was more appropriate for the analysis. Sensitivity analyses, including the leave-one-out method and funnel plots, were conducted to evaluate the robustness of the results and identify potential outliers.

After Bonferroni correction for multiple comparisons, a p-value < 0.001 (0.05/43) was considered indicative of a strong causal relationship, whereas p-values between 0.001 and 0.05 were regarded as evidence of a general association. Reverse causality analyses were performed using MR, MR-PRESSO, and the TwoSampleMR package in R v. 4.3.1 (R Foundation for Statistical Computing, Vienna, Austria). The results were reported in accordance with the Strengthening the Reporting of Observational Studies in Epidemiology–Mendelian Randomization (STROBE-MR) guidelines (Supplementary Table 2).²⁸

Mediation analysis

Mediation analysis is a statistical approach used to examine the mechanisms through which an independent variable (X) influences a dependent variable (Y) via a mediator (M). The objective is to determine whether the effect of X on Y is direct or occurs indirectly through M. This technique is particularly valuable for identifying the underlying pathways that explain relationships between variables. In this study, we applied a simple mediation model in which the independent variables (41 circulating inflammatory factors) influence the mediator (CRP), which in turn affects the dependent variables (sepsis and sepsis-related 28-day mortality).

The effects of the 41 circulating inflammatory factors on sepsis and sepsis-related 28-day mortality can be divided into direct and indirect components. The indirect effect represents the influence of these inflammatory factors on sepsis and 28-day mortality mediated through CRP, whereas the direct effect reflects their influence independent of CRP.²⁹ The proportion of the total effect explained by mediation was estimated by dividing the indirect effect by the overall effect and calculating the 95% confidence interval (95% CI) using the delta method.

Results

Selection of genetic IVs

We screened IVs for CRP, PCT and 41 circulating inflammatory factors and sepsis, respectively. Ultimately,

1,198 SNPs for inflammatory factor, 627 sepsis-associated SNPs, and 1,036 SNPs related to 28-day mortality of sepsis were included. The F-statistics of the genetic IVs that were strongly correlated with inflammatory factors were all greater than 10, indicating a low likelihood of bias influencing the estimates (Supplementary Tables 3,4).

Two-sample MR

Impacts of 41 circulating inflammatory factors, CRP, and PCT on sepsis

This study identified 2 causal correlations between CRP, PCT and 41 circulating inflammatory factors and the risk of sepsis (Fig. 2, Supplementary Table 3). Interleukin 6 (odds ratio (OR) = 0.923, 95% CI: 0.854–0.998; $p = 0.044$), RANTES (OR = 0.926, 95% CI: 0.862–0.994; $p = 0.033$), and macrophage inflammatory protein 1 beta (MIP1 β) (OR = 0.957, 95% CI: 0.919–0.996; $p = 0.032$) were all related to the reduced risk of sepsis. Elevated CRP levels were positively associated with the risk of sepsis (OR = 1.140, 95% CI: 1.055–1.232; $p = 0.001$). Platelet-derived growth factor-BB (PDGFbb) noticeably increased in sepsis (OR = 1.136, CI: 1.003–1.286; $p = 0.044$). According to Cochran's Q test results, the selected SNPs did not exhibit significant heterogeneity ($p > 0.05$; Supplementary Table 5). Similarly, the MR-Egger intercept test indicated no evidence of pleiotropy or outliers ($p > 0.05$; Supplementary Table 5). Finally, the leave-one-out analysis confirmed the robustness of the results (Supplementary Fig. 1–5).

The influence of 41 circulating inflammatory factors, CRP, and PCT on the 28-day mortality of sepsis

The IVW results indicated that CRP, PCT and the 41 circulating inflammatory factors were causally associated with 28-day sepsis-related mortality (Fig. 3; Supplementary Table 4). Genetically predicted CRP was associated with an increased risk of 28-day sepsis mortality (OR = 1.241; 95% CI: 1.034–1.489; $p = 0.020$). Cochran's Q test results showed no significant heterogeneity among the selected SNPs ($p > 0.05$; Supplementary Table 6), and the MR-Egger intercept test revealed no evidence of pleiotropy or outliers ($p > 0.05$; Supplementary Table 6). Finally, the leave-one-out analysis and funnel plot confirmed the stability of the CRP results (Supplementary Fig. 11,12). The Bonferroni-corrected analysis revealed a strong association between CRP levels and the prevalence of sepsis, although this does not imply direct causation (OR = 1.140; 95% CI: 1.055–1.232; $p = 0.001$).

The mediating proportion of CRP in the relation between 41 circulating inflammatory factors and sepsis

We analyzed whether CRP acted as a mediator to strengthen the effects of 41 circulating inflammatory factors on sepsis. It was found that IL-6 and RANTES

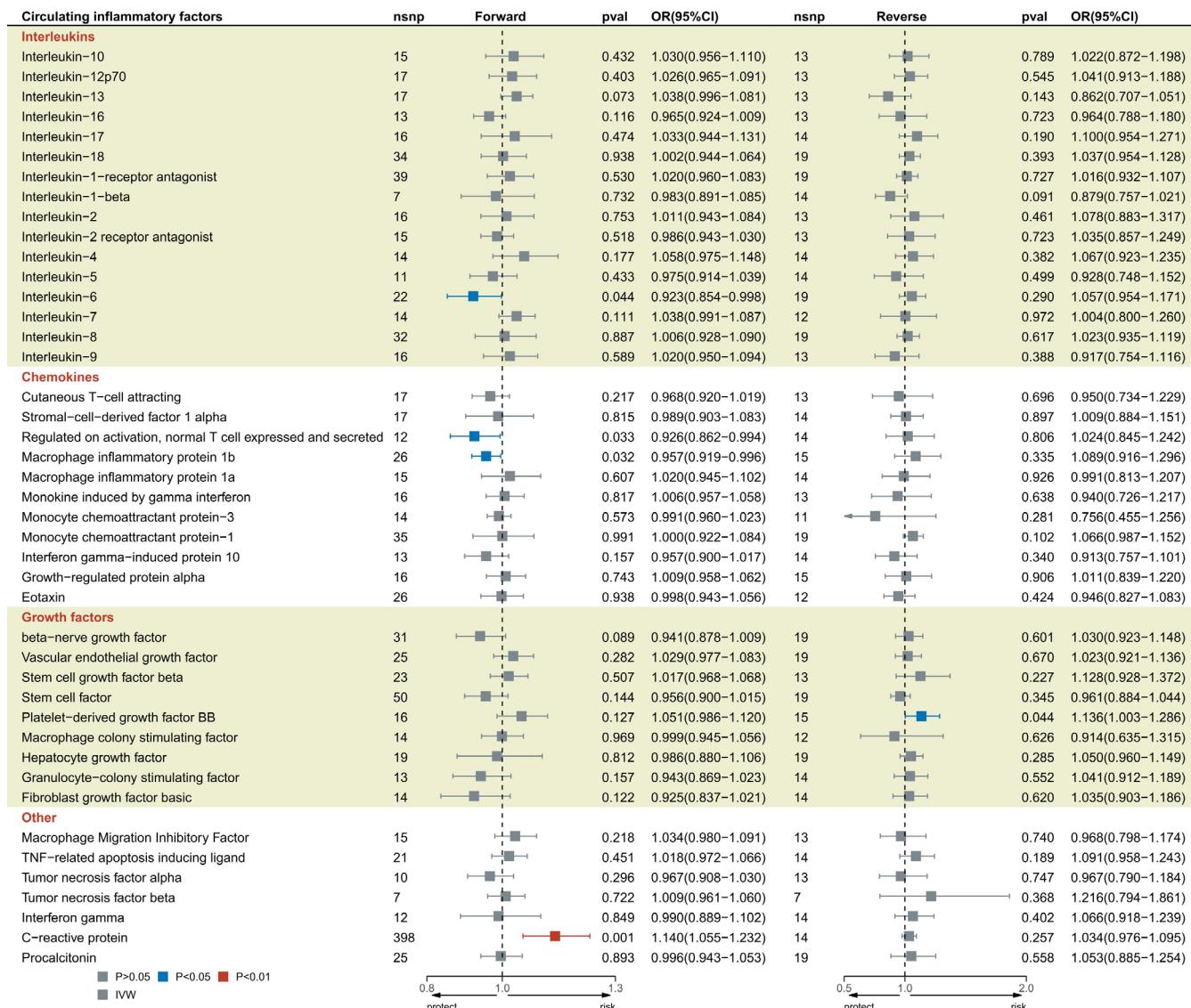


Fig. 2. Forest plot for 41 circulating inflammatory factors, C-reactive protein (CRP) and procalcitonin (PCT) on sepsis

OR – odds ratio; 95% CI – 95% confidence interval; IVW – inverse variance weighted.

were associated with elevated CRP, and in turn, elevated CRP correlated with an enhanced risk of sepsis. As presented in Supplementary Table 7, our research showed that the mediating proportion of CRP in IL-6-based prediction of sepsis was 25.87% (OR = 0.980, 95% CI: 0.961–0.998), and that of CRP in RANTES-based prediction of sepsis was 2.04% (OR = 1.002, 95% CI: 0.991–1.012).

Discussion

This research evaluated the causation between CRP, PCT and 41 circulating inflammatory factors and sepsis/28-day sepsis-associated mortality. Also, mediation analysis was utilized to reveal the mediators of their causal relationships. Important findings were obtained: IL-6, RANTES and MIP1 β were protective factors for sepsis, while CRP

was a risk factor for the onset and 28-day mortality of sepsis. No significant causal relationships were observed between 39 inflammatory factors – including IL-10, PCT and β -NGF – and sepsis or 28-day sepsis-related mortality. Sepsis was identified as the cause of elevated PDGFbb levels. The mediation analysis indicated that the CRP-mediated predictive contribution of IL-6 and RANTES to the onset of sepsis was 25.87% and 2.04%, respectively.

The abnormal production and regulation of inflammatory factors play a crucial role in the pathogenesis of sepsis. Imbalances in these mediators can trigger dysregulated immune responses, leading to an excessive inflammatory cascade. Interleukin 6 is produced rapidly and transiently during infection or trauma, contributing to host defense by promoting acute-phase responses, hematopoiesis and immune activation, thereby playing a key role in the inflammatory process.^{30,31}

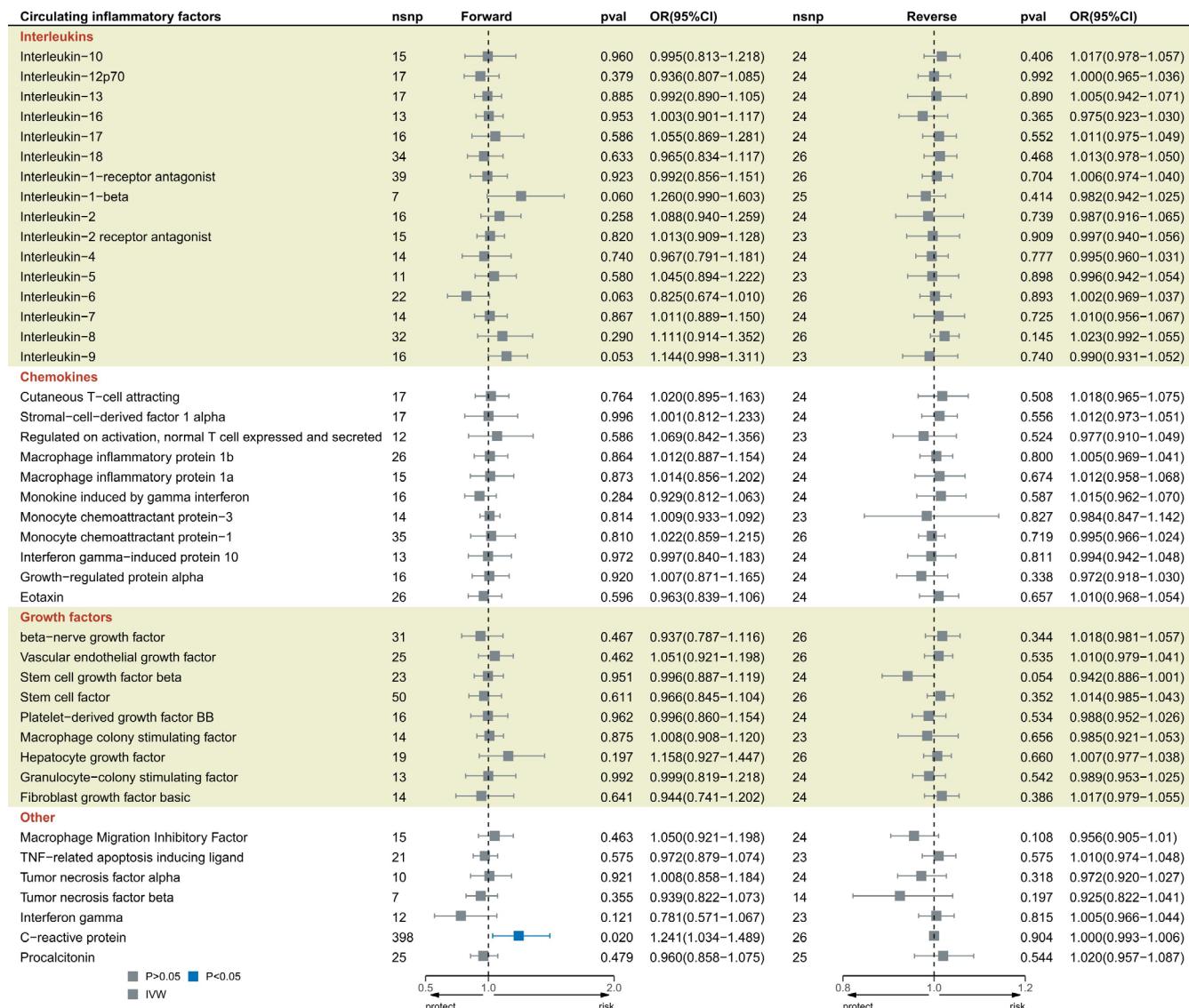


Fig. 3. Forest plot for 41 circulating inflammatory factors, C-reactive protein (CRP) and procalcitonin (PCT) on sepsis (28-day death)

OR – odds ratio; 95% CI – 95% confidence interval; IVW – inverse variance weighted.

Several previous studies have examined the relationship between IL-6 and sepsis. Clinical research has shown that IL-6 levels are elevated in septic patients admitted to the ICU,^{32,33} and that IL-6 is strongly correlated with sepsis severity.³⁴ These findings suggest that IL-6 may serve as a potential biomarker for disease severity in sepsis. Elevated IL-6 levels may also indicate treatment response and prognosis.³⁵ This conclusion is further supported by animal studies.³⁶ Moreover, the combined measurement of TNF- α , IL-1 β and IL-6 has demonstrated strong predictive value for sepsis-associated myocardial injury and is closely related to patient outcomes.³⁷

However, our research findings did not support these conclusions. More studies are necessary in the future to elucidate the causation. While some studies have shown that IL-6 can have protective effects in specific contexts, such as protecting cardiomyocytes from

oxidative stress at the early stage of lipopolysaccharide (LPS)-induced sepsis, it is important to note that literature and experimental results show that IL-6 is a complex marker with both pro-inflammatory and anti-inflammatory roles in sepsis. Given the complex and dual roles of IL-6 in sepsis, clinicians should interpret elevated IL-6 levels with caution.

RANTES is a chemokine involved in regulating cell migration, and its levels progressively increase during infection in patients with sepsis.^{38,39} Elevated concentrations of RANTES have also been observed in plasma samples from cats with sepsis and septic shock.⁴⁰ A small-scale MR analysis previously reported that higher RANTES levels were associated with a reduced risk of sepsis,² indirectly supporting our findings that elevated RANTES concentrations may be protective. RANTES plays an important role in promoting inflammation and activating

other inflammatory mediators such as IL-6. It may serve as an early biomarker of inflammation, aiding in the early diagnosis and monitoring of sepsis. However, similar to CRP, these markers should be interpreted with caution.

Clinical evidence shows no noticeable correlation of MIP1 β with the high risk of sepsis.⁴¹ However, an opposite conclusion was proposed in another prospective and retrospective case-control study that MIP1 β acted as a useful clinical marker in children with sepsis, distinguishing it from other febrile illnesses.⁴² Another prospective clinical study also corroborated that MIP1 β could be applied to distinguish patients with and without sepsis. Our MR analysis illustrated that MIP1 β shared a negative causal association with sepsis and might have a protective impact on sepsis, which warrants verification. Mechanistically, MIP1 β is a member of the chemokine family with chemotactic and pro-inflammatory properties. The release of MIP1 β can activate TCC88 at the site of infection, enhancing the host's resistance to pathogens.⁴³ Future research should further investigate its causal role in sepsis, particularly its influence on inflammatory pathways, and large-scale clinical studies are needed to validate its prognostic significance.

C-reactive protein is an acute-phase protein synthesized in response to cytokine secretion.⁴⁴ Clinical observational studies have demonstrated that CRP possesses the highest diagnostic accuracy for sepsis^{45,46} and serves as one of the most effective prognostic biomarkers for this condition,⁴⁷ making it valuable for predicting treatment outcomes in sepsis.⁴⁸

This research supports our results that CRP is a significant risk factor and acts as a mediator for IL-6 and RANTES to predict 25.87% and 2.04% of sepsis, respectively. Although our study demonstrates a strong link between CRP levels and the prevalence of sepsis, it is important to note that CRP is not an ideal indicator of therapy effectiveness and can be influenced by factors other than infection, such as trauma, surgery, immune system dysregulation, and medications. Given the complexity of sepsis and the multifactorial nature of CRP elevation, further studies are necessary to elucidate the exact role of CRP in the pathogenesis and management of sepsis. This will help in developing more accurate diagnostic tools and therapeutic strategies.

It has been revealed that many inflammatory factors, including IL-1RA, IL-1 β , IL-7, IL-4, IL-9, IL-10, IL-12, IL-13, IL-16, IL-17, and IL-18, are feasible serum biomarkers for sepsis and exert a significant predictive role in the prognosis of this disease.^{2,36,37,41,42,49-52} CAP-1 and TNF- α levels are elevated in patients with sepsis.^{36,49} While TNF- α is a key mediator of the inflammatory response in sepsis, genetic polymorphisms may influence individual susceptibility to its development.⁵³

Studies have shown that sepsis is associated with granulocyte colony-stimulating factor (G-CSF) and monocyte chemoattractant protein-1 (MCP-1) levels, while severe

sepsis-related mortality or sequelae are linked to reduced interferon gamma (IFN- γ) concentrations.⁴¹ TRAIL levels are inversely correlated with sepsis severity,⁵⁴ whereas PCT, IP10 and HGF have been identified as potential serum biomarkers with high diagnostic value and strong prognostic utility in sepsis.^{42,50,55} Serum vascular endothelial growth factor (VEGF) and stromal cell-derived factor 1 (SDF-1) are remarkably correlated with the survival rate of septic patients.^{56,57} Additionally, previous MR analyses have reported a positive causal relationship between beta nerve growth factor (β -NGF) and sepsis.^{2,3} However, in our study, 39 of the circulating inflammatory factors examined did not show significant causal associations.

The reverse MR analysis showed an increase in PDGFbb in sepsis. Previous clinical and preclinical research results supported our finding that the PDGFbb levels are significantly elevated in sepsis.⁵⁸ The mechanism of this correlation may be related to the inhibitory function of PDGFbb in immune cell activation and cytokine production. Platelet-derived growth factor-BB protects against sepsis by reducing the secretion of pro-inflammatory cytokines and chemokines.⁵⁸ This discovery suggests a potential role for PDGFbb in sepsis treatment. While our analysis suggests a potential role for PDGFbb in sepsis treatment, it is important to note that the current understanding of PDGFbb as a therapeutic target in sepsis is still in its early stages. Further studies are necessary to validate the therapeutic potential of PDGFbb and to explore its mechanisms of action in the context of sepsis.

To our knowledge, although previous studies have conducted two-sample MR analyses investigating the causal relationships between 41 circulating inflammatory factors and sepsis, our study offers distinct advantages and provides several novel and suggestive findings. Fang et al. used the MR approach to demonstrate that β -NGF increased the risk of sepsis, whereas RANTES and fibroblast growth factor (FGF) were associated with a reduced risk.² However, their study employed a two-sample unidirectional MR design and was limited by a relatively small GWAS sample size.

Lin et al. conducted a two-sample bidirectional MR study, demonstrating that the predicted circulating RANTES, basic-FGF and β -NGF levels were causally correlated with changes in the risk of sepsis.³ However, these 2 MR analyses did not examine the association between inflammatory factors and sepsis-related mortality, nor did they perform mediation analysis.

Zhi et al. performed a bidirectional MR analysis of circulating inflammatory factors and 28-day sepsis-related mortality, revealing inverse associations between IL-10 and sepsis, as well as between MCP-1 and sepsis-related mortality. In contrast, elevated MIP1 β concentrations showed a positive association with both the incidence and mortality of sepsis.⁵⁹ However, their study did not include mediation analysis. In contrast, our research

employed a bidirectional MR approach using the most recent and comprehensive GWAS datasets to investigate the risk of sepsis and 28-day mortality. Furthermore, we performed mediation analysis to identify potential mediators underlying the observed causal relationships. Notably, although previous MR analyses identified β -NGF and MCP-1 as significant factors, our results showed no significant causal associations between these 2 markers and the risk of sepsis or 28-day sepsis-related mortality. Meanwhile, CRP was identified as a significant predictor of sepsis onset and 28-day mortality, as well as the principal mediator through which IL-6 and RANTES influenced sepsis risk. In addition, PDGFbb emerged as a potential key therapeutic target for sepsis. Heterogeneity and pleiotropy were evaluated, and the results were rigorously tested using multiple sensitivity analyses, confirming the robustness and validity of our findings. C-reactive protein levels rise later than those of IL-6 and, therefore, are not an ideal indicator of therapeutic effectiveness. Although elevated CRP concentrations can indicate the presence of inflammation, they may also be influenced by non-infectious factors such as trauma, surgery or immune dysregulation. Our findings suggest a potential link between IL-6, RANTES, MIP1 β , CRP, PDGFbb, and sepsis severity, but further studies are warranted to elucidate whether these biomarkers play a causative role in the progression of sepsis or if they are merely markers of the underlying disease process.

Limitations

The limitations should also be mentioned. First, because most of the participants recruited in GWAS are from Europe, this population restriction might limit the application of our findings to other populations. Second, although our findings identify the mediators and causal links between CRP, PCT and 41 circulating inflammatory factors and sepsis, the underlying mechanisms require more investigation.

Conclusions

We identified strong causal associations between IL-6, RANTES and MIP1 β and the risk of sepsis. C-reactive protein showed a robust causal relationship with an increased risk of sepsis and was also associated with 28-day sepsis-related mortality. In contrast, 39 inflammatory factors, including IL-10, PCT and β -NGF, showed no significant causal relationships with sepsis risk or 28-day mortality. Platelet-derived growth factor-BB levels were elevated in septic patients. Moreover, CRP acted as a mediating factor in the effects of IL-6 and RANTES on sepsis. These findings may provide valuable insights for the early identification of sepsis and the development of potential therapeutic strategies.

Supplementary data

The supplementary materials are available at <https://doi.org/10.5281/zenodo.15881855>. The package includes the following files:

Supplementary Fig. 1. Leave-one-out sensitivity analysis for the effect of IL-6 on sepsis risk.

Supplementary Fig. 2. Leave-one-out sensitivity analysis for the effect of RANTES on sepsis risk.

Supplementary Fig. 3. Leave-one-out sensitivity analysis for the effect of MIP1 β on sepsis risk.

Supplementary Fig. 4. Leave-one-out sensitivity analysis for the effect of CRP on sepsis risk.

Supplementary Fig. 5. Leave-one-out sensitivity analysis for the effect of sepsis on PDGF-BB levels.

Supplementary Fig. 6. Funnel plot assessing asymmetry in the MR estimates for the effect of IL-6 on sepsis risk.

Supplementary Fig. 7. Funnel plot assessing asymmetry in the MR estimates for the effect of RANTES on sepsis risk.

Supplementary Fig. 8. Funnel plot assessing asymmetry in the MR estimates for the effect of MIP1 β on sepsis risk.

Supplementary Fig. 9. Funnel plot assessing asymmetry in the MR estimates for the effect of CRP on sepsis risk.

Supplementary Fig. 10. Funnel plot assessing asymmetry in the MR estimates for the effect of sepsis on PDGFbb levels.

Supplementary Fig. 11. Leave-one-out sensitivity analysis for the effect of CRP on sepsis (28-day death) risk.

Supplementary Fig. 12. Funnel plot assessing asymmetry in the MR estimates for the effect of CRP on sepsis (28-day death) risk.

Supplementary Table 1. Sources and details of the GWAS summary statistics used for CRP, PCT and 41 circulating inflammatory factors.

Supplementary Table 2. Analytical methods used for MR and mediation analysis.

Supplementary Table 3. Bidirectional MR estimating causal effects between circulating inflammatory factors and sepsis risk.

Supplementary Table 4. Bidirectional MR results estimating causal effects between circulating inflammatory factors and sepsis (28-day death).

Supplementary Table 5. Heterogeneity and horizontal pleiotropy assessments for MR analyses of circulating inflammatory factors on sepsis risk.

Supplementary Table 6. Heterogeneity and horizontal pleiotropy assessments for MR analyses of circulating inflammatory factors on sepsis (28-day death).

Supplementary Table 7. Two-step MR analysis quantifying mediation effects of IL-6 and RANTES on the relationship between CRP and sepsis risk.

Data Availability Statement

Summaries of sepsis and circulating inflammatory factors were acquired from the IEU OpenGWAS Project (<https://gwas.mrcieu.ac.uk/>) and OpenGWAS databases.

Consent for publication

Not applicable.

Use of AI and AI-assisted technologies

Not applicable.

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References

1. Singer M, Deutschman CS, Seymour CW, et al. The Third International Consensus Definitions for Sepsis and Septic Shock (Sepsis-3). *JAMA*. 2016;315(8):801. doi:10.1001/jama.2016.0287
2. Fang W, Chai C, Lu J. The causal effects of circulating cytokines on sepsis: A Mendelian randomization study. *PeerJ*. 2024;12:e16860. doi:10.7717/peerj.16860
3. Lin S, Mao X, He W. Causal association of circulating cytokines with sepsis: A Mendelian randomization study. *Front Immunol*. 2023;14:1281845. doi:10.3389/fimmu.2023.1281845
4. Rudd KE, Johnson SC, Agesa KM, et al. Global, regional, and national sepsis incidence and mortality, 1990–2017: Analysis for the Global Burden of Disease Study. *Lancet*. 2020;395(10219):200–211. doi:10.1016/S0140-6736(19)32989-7
5. García-Rodríguez JF, Mariño-Callejo A. The factors associated with the trend in incidence of bacteraemia and associated mortality over 30 years. *BMC Infect Dis*. 2023;23(1):69. doi:10.1186/s12879-023-08018-0
6. Fleischmann C, Scherag A, Adhikari NKJ, et al. Assessment of global incidence and mortality of hospital-treated sepsis: Current estimates and limitations. *Am J Respir Crit Care Med*. 2016;193(3):259–272. doi:10.1164/rccm.201504-0781OC
7. Li A, Ling L, Qin H, et al. Epidemiology, management, and outcomes of sepsis in ICUs among countries of differing national wealth across Asia. *Am J Respir Crit Care Med*. 2022;206(9):1107–1116. doi:10.1164/rccm.202112-2743OC
8. Kiya GT, Mekonnen Z, Melaku T, et al. Prevalence and mortality rate of sepsis among adults admitted to hospitals in sub-Saharan Africa: A systematic review and meta-analysis. *J Hosp Infect*. 2024;144:1–13. doi:10.1016/j.jhin.2023.11.012
9. Ma Z, Jiang Z, Li H, et al. Prevalence, early predictors, and outcomes of sepsis in neurocritical illnesses: A prospective cohort study. *Am J Infect Control*. 2024;52(7):827–833. doi:10.1016/j.ajic.2024.01.017
10. Yang X, Man MY, Heng H, et al. Molecular epidemiology and clinical impact of *Klebsiella* spp. causing bloodstream infections in Hong Kong. *eBioMedicine*. 2024;101:104998. doi:10.1016/j.ebiom.2024.104998
11. Yang M, Liu Z, Xu Z, et al. Inhibition of Toll-like receptor 4 pathway by TAK242 protects the liver in sepsis [in Chinese]. *Zhonghua Wei Zhong Bing Ji Jiu Yi Xue*. 2022;34(8):814–818. doi:10.3760/cma.j.cn121430-20220420-00395
12. Liu J, Li J, Tian P, et al. H2S attenuates sepsis-induced cardiac dysfunction via a PI3K/Akt-dependent mechanism. *Exp Ther Med*. 2019;17(5):4064–4072. doi:10.3892/etm.2019.7440
13. Ma F, Liu F, Ding L, et al. Anti-inflammatory effects of curcumin are associated with downregulating microRNA-155 in LPS-treated macrophages and mice. *Pharm Biol*. 2017;55(1):1263–1273. doi:10.1080/13880209.2017.1297838
14. Liu Z, Gao J, Ban Y, et al. Synergistic effect of paeoniflorin combined with luteolin in alleviating lipopolysaccharides-induced acute lung injury. *J Ethnopharmacol*. 2024;327:118022. doi:10.1016/j.jep.2024.118022
15. Ling H, Chen M, Dai J, Zhong H, Chen R, Shi F. Evaluation of qSOFA combined with inflammatory mediators for diagnosing sepsis and predicting mortality among emergency department. *Clin Chim Acta*. 2023;544:117352. doi:10.1016/j.cca.2023.117352
16. Zhang GM, Gu YY. Diagnostic value of procalcitonin, C-reactive protein-to-lymphocyte ratio (CLR), C-reactive protein and neutrophil-to-lymphocyte ratio (NLR) for predicting patients with bacteraemia in the intensive care unit. *J Crit Care*. 2024;81:154538. doi:10.1016/j.jcrc.2024.154538
17. Fahmy EM, Kamel NM, Abdelsadik A, et al. Assessment of interleukin-27 and chemokine RANTES as biomarkers for early onset neonatal sepsis. *Egypt J Immunol*. 2020;27(1):9–18. PMID:33180383.
18. Emdin CA, Khera AV, Kathiresan S. Mendelian randomization. *JAMA*. 2017;318(19):1925. doi:10.1001/jama.2017.17219
19. Walker VM, Zheng J, Gaunt TR, Smith GD. Phenotypic causal inference using genome-wide association study data: Mendelian randomization and beyond. *Annu Rev Biomed Data Sci*. 2022;5(1):1–17. doi:10.1146/annurev-biodatasci-122120-024910
20. Ahola-Olli AV, Würtz P, Havulinna AS, et al. Genome-wide association study identifies 27 loci influencing concentrations of circulating cytokines and growth factors. *Am J Hum Genet*. 2017;100(1):40–50. doi:10.1016/j.ajhg.2016.11.007
21. Kaloja M, Corbin LJ, Tan VY, et al. The role of inflammatory cytokines as intermediates in the pathway from increased adiposity to disease. *Obesity*. 2021;29(2):428–437. doi:10.1002/oby.23060
22. FolkerSEN L, Gustafsson S, Wang Q, et al. Genomic and drug target evaluation of 90 cardiovascular proteins in 30,931 individuals. *Nat Metab*. 2020;2(10):1135–1148. doi:10.1038/s42255-020-00287-2
23. Hillary RF, Trejo-Banos D, Kousathanas A, et al. Multi-method genome- and epigenome-wide studies of inflammatory protein levels in healthy older adults. *Genome Med*. 2020;12(1):60. doi:10.1186/s13073-020-00754-1
24. Said S, Pazoki R, Karhunen V, et al. Genetic analysis of over half a million people characterises C-reactive protein loci. *Nat Commun*. 2022;13(1):2198. doi:10.1038/s41467-022-29650-5
25. Sun BB, Maranville JC, Peters JE, et al. Genomic atlas of the human plasma proteome. *Nature*. 2018;558(7708):73–79. doi:10.1038/s41586-018-0175-2
26. Hamilton FW, Thomas M, Arnold D, et al. Therapeutic potential of IL6R blockade for the treatment of sepsis and sepsis-related death: A Mendelian randomization study. *PLoS Med*. 2023;20(1):e1004174. doi:10.1371/journal.pmed.1004174
27. Wang E, Zhao H, Zhao D, Li L, Du L. Functional prediction of chronic kidney disease susceptibility gene PRKAG2 by comprehensively bioinformatics analysis. *Front Genet*. 2018;9:573. doi:10.3389/fgene.2018.00573
28. Skrvankova VW, Richmond RC, Woolf BAR, et al. Strengthening the Reporting of Observational Studies in Epidemiology Using Mendelian Randomization: The STROBE-MR Statement. *JAMA*. 2021;326(16):1614. doi:10.1001/jama.2021.18236
29. Carter AR, Sanderson E, Hammerton G, et al. Mendelian randomisation for mediation analysis: Current methods and challenges for implementation. *Eur J Epidemiol*. 2021;36(5):465–478. doi:10.1007/s10654-021-00757-1
30. Tanaka T, Narazaki M, Kishimoto T. IL-6 in inflammation, immunity, and disease. *Cold Spring Harb Perspect Biol*. 2014;6(10):a016295-a016295. doi:10.1101/cshperspect.a016295
31. Unver N, McAllister F. IL-6 family cytokines: Key inflammatory mediators as biomarkers and potential therapeutic targets. *Cytokine Growth Factor Rev*. 2018;41:10–17. doi:10.1016/j.cytogfr.2018.04.004
32. Nader D, Abdelmaseeh M, Al Sharif R, Samir N. Serum level of microRNA 15a and interleukin 6 as biomarkers for sepsis in critically ill patients. *Egypt J Immunol*. 2023;30(3):162–170. PMID:37440665.
33. Jiang S, Shi D, Bai L, Niu T, Kang R, Liu Y. Inhibition of interleukin-6 trans-signaling improves survival and prevents cognitive impairment in a mouse model of sepsis. *Int Immunopharmacol*. 2023;119:110169. doi:10.1016/j.intimp.2023.110169
34. Cao Z, Wu M, Li Y, et al. Predictive effect of combined procalcitonin, interleukin-6 and antithrombin III on the severity and prognosis of patients with sepsis [in Chinese]. *Zhonghua Wei Zhong Bing Ji Jiu Yi Xue*. 2023;35(10):1033–1038. doi:10.3760/cma.j.cn121430-20221114-00981

35. Palalioğlu B, Erdoğan S, Atay G, Tugrul H, Özer Ö. Diagnostic and prognostic value of pentraxin 3, interleukin-6, CRP, and procalcitonin levels in patients with sepsis and septic shock. *Niger J Clin Pract.* 2024;27(3):317–324. doi:10.4103/njcp.njcp_615_23

36. Cao YZ, Tu YY, Chen X, Wang BL, Zhong YX, Liu MH. Protective effect of ulinastatin against murine models of sepsis: Inhibition of TNF- α and IL-6 and augmentation of IL-10 and IL-13. *Exp Toxicol Pathol.* 2012;64(6):543–547. doi:10.1016/j.etp.2010.11.011

37. Deng P, Tang N, Li L, Zou G, Xu Y, Liu Z. Diagnostic value of combined detection of IL-1 β , IL-6, and TNF- α for sepsis-induced cardiomyopathy. *Med Clin (Barc).* 2022;158(9):413–417. doi:10.1016/j.medcli.2021.04.025

38. Stojewska M, Wąsek-Buko M, Jakub B, et al. Evaluation of serum chemokine RANTES concentration as a biomarker in the diagnosis of early-onset severe infections in neonates. *Postepy Hig Med Dosw.* 2016;70:272–279. doi:10.5604/17322693.1198990

39. Cavalcanti NV, Torres LC, Da Matta MC, et al. Chemokine patterns in children with acute bacterial infections. *Scand J Immunol.* 2016;84(6):338–343. doi:10.1111/sji.12492

40. Troia R, Mascalzoni G, Agnoli C, Lalonde-Paul D, Giunti M, Goggs R. Cytokine and chemokine profiling in cats with sepsis and septic shock. *Front Vet Sci.* 2020;7:305. doi:10.3389/fvets.2020.00305

41. Leal YA, Álvarez-Nemegyei J, Lavadores-May AI, Girón-Carrillo JL, Cedillo-Rivera R, Velazquez JR. Cytokine profile as diagnostic and prognostic factor in neonatal sepsis. *J Matern Fetal Neonatal Med.* 2019;32(17):2830–2836. doi:10.1080/14767058.2018.1449828

42. Frimpong A, Owusu EDA, Amponsah JA, et al. Cytokines as potential biomarkers for differential diagnosis of sepsis and other non-septic disease conditions. *Front Cell Infect Microbiol.* 2022;12:901433. doi:10.3389/fcimb.2022.901433

43. Naghavian R, Faigle W, Oldrati P, et al. Microbial peptides activate tumour-infiltrating lymphocytes in glioblastoma. *Nature.* 2023;617(7962):807–817. doi:10.1038/s41586-023-06081-w

44. Plebani M. Why C-reactive protein is one of the most requested tests in clinical laboratories? *Clin Chem Lab Med.* 2023;61(9):1540–1545. doi:10.1515/cclm-2023-0086

45. Goyal M, Mascarenhas D, Rr P, Haribalakrishna A. Diagnostic accuracy of point-of-care testing of C-reactive protein, interleukin-6, and procalcitonin in neonates with clinically suspected sepsis: A prospective observational study. *Med Princ Pract.* 2024;33(3):291–298. doi:10.1159/000536678

46. Cui N, Zhang YY, Sun T, Lv XW, Dong XM, Chen N. Utilizing procalcitonin, C-reactive protein, and serum amyloid A in combination for diagnosing sepsis due to urinary tract infection. *Int Urol Nephrol.* 2024;56(7):2141–2146. doi:10.1007/s11255-024-03959-0

47. Tocu G, Mihailov R, Serban C, Stefanescu BI, Tutunaru D,irescu D. The contribution of procalcitonin, C-reactive protein and interleukin-6 in the diagnosis and prognosis of surgical sepsis: An observational and statistical study. *J Multidiscip Healthc.* 2023;16:2351–2359. doi:10.2147/JMDH.S422359

48. Jimoh A, Bolaji O, Adelekan A, et al. Clinical utility of procalcitonin and C-reactive protein in the management of neonatal sepsis in a resource-limited Nigerian hospital. *Niger J Clin Pract.* 2023;26(12):1895–1901. doi:10.4103/njcp.njcp_397_23

49. Li G, Yang Z, Yang C, et al. Single-cell RNA sequencing reveals cell-cell communication and potential biomarker in sepsis and septic shock patients. *Int Immunopharmacol.* 2024;132:111938. doi:10.1016/j.intimp.2024.111938

50. Zhang W, Wang W, Hou W, et al. The diagnostic utility of IL-10, IL-17, and PCT in patients with sepsis infection. *Front Public Health.* 2022;10:923457. doi:10.3389/fpubh.2022.923457

51. Eidl MV, Nunes FB, Pedrazza L, et al. Biochemical and inflammatory aspects in patients with severe sepsis and septic shock: The predictive role of IL-18 in mortality. *Clin Chim Acta.* 2016;453:100–106. doi:10.1016/j.cca.2015.12.009

52. Froeschle GM, Bedke T, Boettcher M, Huber S, Singer D, Ebenebe CU. T cell cytokines in the diagnostic of early-onset sepsis. *Pediatr Res.* 2021;90(1):191–196. doi:10.1038/s41390-020-01248-x

53. Georgescu AM, Banescu C, Azamfirei R, et al. Evaluation of TNF- α genetic polymorphisms as predictors for sepsis susceptibility and progression. *BMC Infect Dis.* 2020;20(1):221. doi:10.1186/s12879-020-4910-6

54. Yoo H, Lee JY, Park J, Yang JH, Suh GY, Jeon K. Association of plasma level of TNF-related apoptosis-inducing ligand with severity and outcome of sepsis. *J Clin Med.* 2020;9(6):1661. doi:10.3390/jcm9061661

55. Peng F, Liang C, Chang W, et al. Prognostic significance of plasma hepatocyte growth factor in sepsis. *J Intensive Care Med.* 2022;37(3):352–358. doi:10.1177/0885066621993423

56. Patry C, Stamm D, Betzen C, et al. CXCR-4 expression by circulating endothelial progenitor cells and SDF-1 serum levels are elevated in septic patients. *J Inflamm (Lond).* 2018;15(1):10. doi:10.1186/s12950-018-0186-7

57. Siavashi V, Asadian S, Taheri-Asl M, Keshavarz S, Zamani-Ahmadmahmudi M, Nassiri SM. Endothelial progenitor cell mobilization in preterm infants with sepsis is associated with improved survival. *J Cell Biochem.* 2017;118(10):3299–3307. doi:10.1002/jcb.25981

58. Wang M, Wei J, Shang F, Zang K, Ji T. Platelet-derived growth factor B attenuates lethal sepsis through inhibition of inflammatory responses. *Int Immunopharmacol.* 2019;75:105792. doi:10.1016/j.intimp.2019.105792

59. Zhi F, Ma JW, Ji DD, Bao J, Li QQ. Causal associations between circulating cytokines and risk of sepsis and related outcomes: A two-sample Mendelian randomization study. *Front Immunol.* 2024;15:1336586. doi:10.3389/fimmu.2024.1336586