**GP6 promotes the development of cerebral ischemic stroke induced by atherosclerosis via the FYN-PKA-pPTK2/FAK1 signaling pathway**

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A – research concept and design; B – collection and/or assembly of data; C – data analysis and interpretation;  
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**Abstract**

**Background.** Cerebrovascular disease is currently a serious threat to human health and life, commonly including cerebral infarction, cerebral hemorrhage and transient cerebral ischemia, among others.

**Objectives.** To explore the role and molecular mechanism of GP6 in the development of cerebral ischemic stroke (CIS) induced by atherosclerosis (AS).

**Materials and methods.** Forty-five male New Zealand white rabbits were randomly divided into 3 groups: the control, CIS model and anti-GP6 group. Carotid artery tissues and blood of the white rabbits were collected for analysis. Hematoxylin and eosin (H&E) staining was used to analyze the pathological characteristics of vascular endothelial injury. Flow cytometry (FCM) was performed to analyze the content of Th1 and Th17 in blood. Immunohistochemistry was used to analyze the distribution and relative expression of FCER1G, ITGA2 and GP6 proteins in the carotid artery and cerebrovascular tissues. Western blot was applied to determine the protein expression of GP6, FYN, PKA, pPTK2, and pFAK1 in carotid artery tissues of the rabbits.

**Results.** In the CIS model group, there was lymphocyte infiltration, fibrous tissue formation, and the formation of thrombus and lipid plaques. In the anti-GP6 group, scattered thin plaques were observed, and no obvious foam cell deposition was observed. The Th1 and Th17 content was significantly decreased in the CIS model group compared to the control and anti-GP6 group. The relative expression of FCER1G, ITGA2 and GP6 in the CIS model group was significantly higher compared to those in the control group and anti-GP6 group. The protein expression of GP6, FYN, PKA, pPTK2, and pFAK1 in the CIS model group were markedly higher compared to those in the control group and anti-GP6 group.

**Conclusions.** GP6 can promote the development of CIS by activating the FYN-PKA-pPTK2/FAK1 signaling pathway.

**Key words:** CIS, AS, GP6, thrombus, vascular endothelial cells
Background

Cerebrovascular disease is currently a serious threat to human health and life, the most common being cerebral infarction, cerebral hemorrhage and transient cerebral ischemia, among others. It mainly occurs in middle-aged and elderly people, but the age of onset has shown a trend of decreasing, often occurring in people 30–50 years old. The disease often leads to cognitive disorders or disorders affecting the physical activity of patients. Many patients are unable to take care of themselves after onset and quality of life is seriously reduced, resulting in them becoming a heavy burden to their families and society. At present, there is no radically effective cure for the disease, and the prognosis is often closely related to the initial severity of the disease. Even if patients survive, most of them are accompanied by lifelong sequelae, and cerebrovascular diseases can occur repeatedly, causing far more pain to the patients and their families than malignant tumors and cardiovascular diseases. Therefore, how to prevent and treat cerebrovascular disease more effectively is the most serious problem that neurology faces presently. Cerebral ischemic stroke (CIS) is ischemic necrosis or softening of localized brain tissue due to cerebral blood circulation disorder, ischemia or hypoxia. It is the most common type of cerebrovascular disease, accounting for about 70% of all acute cerebrovascular diseases. About 30–40% of CIS is caused by carotid atherosclerotic stenosis.

Extracellular matrix (ECM) contact exposed at the site of platelet and vascular injury is the first line of defense for repairing damaged tissue and stopping bleeding. Collagen, one of the macromolecular components of ECM, can not only adhere to platelets through direct and indirect channels but also cause platelet aggregation and expression of pro-coagulant activity. The interaction between platelets and collagen in arteries or damaged vessels with high shear stress is particularly important. Current research suggests that platelets have 2 types of collagen receptors on their surfaces, α2β1 and glycoprotein 6 (GP6). It is a glycoprotein receptor that acts on collagen and is located on the long arm of chromosome 19 (19q13.4). It contains 8 exons and has a molecular weight of 62 kDa. In humans, GP6 is encoded by the GP6 gene. GP6 mediates the initial adhesion of platelets to collagen, generates signal transduction, improves the binding affinity of integrin receptors, and causes platelet aggregation, platelet release and thrombosis. When serum GP6 concentration is increased, it can promote the initial adhesion of collagen and platelets, improving the affinity of the integrin receptor, which induces platelet aggregation and thrombosis. Kubisz et al. found that a variation of the GP6 gene was related to platelet aggregation. Sokol et al. found that a GP6 gene polymorphism may be related to the heightened aggregation ability of platelets. Despite the complex composition of the ECM, the adhesion of platelets and collagen plays an important role in initiating hemostasis and thrombus in the body. Inhibition of GP6 function can significantly inhibit collagen-induced platelet adhesion, aggregation and platelet thrombosis under high shear stress in vitro. Therefore, we believe that atherosclerotic CIS is closely related to the structural and functional integrity of GP6, but the mechanism of action of GP6 in atherosclerosis (AS) has rarely been studied.

Objectives

In this paper, we wanted to investigate the mechanism of action of GP6 in AS. Helper T (Th) cell content, Th1 and Th17, was significantly decreased in the CIS model group compared to the control group, while having no marked differences compared to the anti-GP6 group. Immunohistochemistry showed that the proteins FCER1G, ITGA2 and GP6 were all distributed in the cell membrane. GP6 can promote the formation of cerebral ischemic stroke via up-regulating the expression of FCER1G, ITGA2, FYN, PKA, pPTK2, and pFAK1. This may be a new target for treating cerebral ischemic stroke.

Materials and methods

Model establishment and grouping

Forty-five male New Zealand white rabbits (~16–18 weeks old, weight: ~1.5–2.0 kg) were obtained from the Naval Medical Institute Animal Center (Shanghai, China). They were randomly divided into 3 groups (n = 15 in each group): the control group, CIS group and anti-GP6 group. In the CIS group and the anti-GP6 group, rabbits were first fed with a specific high-fat diet for 8 weeks, and then 3% hydrogen peroxide solution was applied to perfuse the bilateral common carotid arteries of the rabbits, causing vascular endothelial oxidative stress injury. In the anti-GP6 group, the rabbits were given anti-GP6 antibody via the tail vein. Carotid artery tissues of the white rabbits were collected and rapidly stored in −80°C liquid nitrogen for further use.

In this study, atherosclerotic stenosis injury rabbit models were constructed and separated into 3 groups: the control group, CIS group and anti-GP6 group. Hematoxylin and eosin (H&E) staining was used to analyze the pathological features of vascular endothelial injury. Flow cytometry (FCM) was used to determine the concentration of Th1 and Th17 cells in blood samples. The expressions of GP6, FYN, PKA, pPTK2, and FAK1 were also assessed.

The animal experiments were approved by the Ethics Committee of Xinhua Hospital affiliated with Shanghai Jiaotong University School of Medicine (Shanghai, China).
Hematoxylin and eosin staining

After the rabbits were sacrificed by the cervical dislocation method, hippocampal tissue was taken from their brains. The tissues were fixed with 4% formaldehyde for 24 h, embedded in paraffin and sectioned into 5 μm-thick sections. Histopathological changes were observed under a light microscope (Olympus Corp., Tokyo, Japan) after H&E staining.

Flow cytometry

Cells were digested with trypsin, washed twice with phosphate-buffered saline (PBS) and collected into a centrifuge tube. Binding buffer was added to prepare cell suspensions with a final concentration of 1 × 10⁶/mL. Annexin V-FITC kit (BioVision, Milpitas, USA) instructions were used for labeling, which was performed according to the instructions of Annexin V-FITC kit. Annexin V was added for staining at room temperature in the dark for 15 min.

Western blot

Cells were lysed with cell lysis buffer on ice for 30 min and then centrifuged at 12,000 rpm for 25 min. Supernatant was collected and the Bradford method was used for protein quantification. Protein in the amount of 25 μg was loaded for sodium dodecyl sulphate–polyacrylamide gel electrophoresis (SDS-PAGE). The proteins were transferred to a polyvinylidene difluoride (PVDF) membrane and sealed with 5% skim milk at room temperature for 1 h; then, primary antibody was added and the membrane was incubated overnight at 4°C. The next day, the secondary antibody was added and incubated at room temperature for 1 h, and then, electrochemiluminescence (ECL) hypersensitive luminescence solution was added for developing.

Immunohistochemistry

Tissues were dewaxed and endogenous oxidase was inactivated with 3% hydrogen peroxide. After washing in PBS 3 times, the antigen was retrieved by microwave heating. The slices were placed in sodium citrate buffer with pH 6.0, heated to boiling in a microwave and then cooled. The slices were again microwaved in sodium citrate buffer with pH 6.0 and then washed with PBS. After cleaning, sections were removed and sealed for 10 min. Then, mouse anti-human FCER1G, ITGA2 and GP6 monoclonal antibodies (1:150) were added and incubated at 4°C for 12 h. After washing, biotin-labeled secondary antibodies were added and incubated at 4°C for 20 min each. After incubation and washing, the samples were washed again and immersed in DAB color developing solution for 5 min. Then, the samples were washed, re-dyed, dehydrated, sealed and observed under an optical microscope.

Statistical analyses

IBM SPSS v. 19.0 software (IBM Corp., Armonk, USA) was used for statistical analysis. The results were expressed as mean values ± standard deviation (SD). Differences between groups were compared using one-way analysis of variance (ANOVA) tests followed by Bonferroni post hoc test. The assumption for ANOVA was verified with the Brown–Forsythe test. A value of p < 0.05 was considered statistically significant.

Results

Anti-GP6 inhibits vascular endothelial injury

After the successful establishment of the evaluation models, the rabbits were anesthetized with isopentane and sacrificed. Carotid artery tissue and cerebrovascular tissues were collected from the white rabbits, and H&E staining was used to analyze the pathological characteristics of the vascular endothelial injury. As shown in Fig. 1, in the control group, the smooth muscle cells were arranged neatly, tightly and orderly, the endothelial layer was continuous and there was no lipid deposition in the subcutaneous tissue. The structure of each layer was clear. In the CIS group, the endothelium was discontinuous, the intimal hyperplasia was serious, the tube wall was full of plaques, a large number of foam cells could be seen, the membranes in the middle and outside were irregularly thickened, and foam cell infiltration could be seen. In the anti-GP6 group, endothelial cells were continuous, lipid deposition was observed under the intima, scattered thin plaques were observed, and no obvious foam cell deposition was observed. The thickness of the membrane was the same as that of the control group membrane, which was thinner than the CIS model group membrane.

Content of Th1 and Th17

The content of Th1 and Th17 in blood was analyzed with FCM and the results are shown in Fig. 2. The Th1 (p < 0.01, degrees of freedom (df) = 2, F = 337.7) and Th17 (p < 0.01, df = 2, F = 150.6) content was significantly decreased in the CIS model group compared to the control group, while there was no marked difference compared to the anti-GP6 group.

Results of immunohistochemistry

Immunohistochemistry was used to analyze the distribution and relative expression of FCER1G, ITGA2 and GP6 proteins in the carotid artery and cerebrovascular tissues of white rabbits. The proteins FCER1G, ITGA2 and GP6 were all distributed in the cell membrane.
Y. Gu, Y. Wu, L. Chen. GP6 promotes the development of CIS (Fig. 3). The relative expression of FCER1G, ITGA2 and GP6 in the CIS model group was significantly higher compared to the control group and anti-GP6 group. The relative expression of FCER1G, ITGA2 and GP6 did not significantly differ between the control group and anti-GP6 group.

Expression of GP6, FYN, PKA, pPTK2, and pFAK1

Western blot was applied to determine the protein expression of GP6, FYN, PKA, pPTK2, and pFAK1 in carotid artery tissues of rabbits from the control, CIS model and anti-GP6 group. The protein expression of GP6 (p < 0.001, df = 2, F = 392.9), FYN (p < 0.001, df = 2, F = 380.3), PKA (p < 0.001, df = 2, F = 304.3), pPTK2 (p < 0.001, df = 2, F = 369.9), and pFAK1 (p < 0.001, df = 2, F = 562.8) in the CIS model group were markedly higher compared to those in the control group and anti-GP6 group (Fig. 4). Moreover, the expression of GP6, FYN, PKA, pPTK2, and pFAK1 were significantly upregulated in the anti-GP6 group compared with the control group.

Discussion

With a remarkable rise in people’s living standards and an accelerated pace of life, the onset of CIS is observed earlier than before and the incidence rate increases year after year. In particular, progressive CIS has a poor prognosis and high morbidity and mortality, which poses a serious threat to people’s health and life safety, and burdens patients and families. Carotid AS is an important risk factor for CIS and is closely related to the occurrence, development and recurrence of cerebral infarction. To date, surgery
Fig. 3. Immunofluorescent staining of FCER1G, ITGA2 and GP6 proteins in the carotid artery and cerebrovascular tissues of white rabbits in the control, cerebral ischemic stroke (CIS) model and anti-GP6 groups. The proteins FCER1G, ITGA2 and GP6 were all distributed in the cell membrane. The relative expression of FCER1G, ITGA2 and GP6 in the CIS model group was significantly higher compared to the control group and anti-GP6 group.

Fig. 4. The protein expression of GP6, FYN, PKA, pPTK2, and pFAK1 in carotid artery tissues of rabbits in the control, cerebral ischemic stroke (CIS) model and anti-GP6 groups measured with western blot assay. The protein expression of GP6, FYN, PKA, pPTK2, and pFAK1 in the CIS model group was markedly higher compared to the control group and anti-GP6 group.

*p < 0.001 or *p < 0.01, indicating the significant difference using one-way analysis of variance (ANOVA) followed by Bonferroni post hoc test.
or endovascular treatment for patients with recent ischemic diseases has been dependent on the degree of narrowing of the responsible vessel. For many years, the extent of artery stenosis has been regarded as the most reliable index to predict the risk of stroke in patients with AS. Thromboembolism derived from atherosclerotic plaque is considered to be the main pathogenesis of CIS in most atherosclerotic patients. Plaque rupture can lead to platelet aggregation, local thrombosis or thromboembolism. Plaque itself can also cause thromboembolism. Protein function analysis revealed that GP6 is a platelet glycoprotein, which plays an important role in platelet coagulation, thrombosis and arterial embolization. The GP6 signaling pathway mainly activates FYN and PKA, phosphorylates PTK2/FAK1 and promotes platelet adhesion to damaged vascular endothelium.

Vascular endothelial cells have anticoagulant and fibrinolytic properties and can influence the chemotaxis and adhesion properties of cell membranes. Under normal circumstances, vascular endothelial cells cover the inner surface of the entire vascular system, providing an anticoagulant interface that prevents platelets and other blood cells from adhering to the subcutaneous tissue and preventing activation of the clotting response. Endocrine dysfunction caused by endothelial cell injury plays a key role in the pathophysiological mechanism of the early formation and development of AS. In this study, male New Zealand white rabbits were randomly divided into 3 groups: control, CIS model and anti-GP6 group. Results of H&E staining analysis of the vascular endothelial injury showed that in the CIS model group, the endothelium was discontinuous, the intimal hyperplasia was serious, the tube wall was full of plaques, a large number of foam cells could be seen, the membranes in the middle and outside were irregularly thickened, and foam cell infiltration could be seen. However, the injury degree was reduced in the anti-GP6 group compared to the CIS model group. The relative expression of H&E staining analysis of the vascular endothelial in question is that the expression of FYN and PKA, phosphorylates PTK2/FAK1 and promotes platelet adhesion to damaged vascular endothelium.

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The role of Th17 cells in disease is mainly to promote defense against organ-specific autoimmune diseases and chronic infections. Results of animal experiments carried out by Davenport et al. showed that Th1 and Th2 immune responses were involved in the formation of atherosclerotic lesions. Kim et al. assessed 124 patients with chest pain who underwent coronary angiography and found the expression of Th1 and Th17 cells in patients with stable angina pectoris was significantly increased, which indicated that Th1 and Th17 cells are associated with the development of AS. Interleukin 17 (IL-17) is a major effector of Th17 cells and an early promoter of T-cell-induced inflammatory response, which can amplify the inflammatory response. Immunohistochemistry results in our study showed the proteins FCER1G, ITGA2 and GP6 were all distributed in the cell membrane. The relative expression of FCER1G, ITGA2 and GP6 in the CIS model group was significantly higher compared to the control group and anti-GP6 group. The FCER1G gene is located on chromosome 1q23 (1:161185024-161190489) and encodes an 86-amino acid protein. ITGA2 is a protein-coding gene. Diseases associated with ITGA2 include anus disease and thrombocytopenia. Results from western blotting indicated that the protein expression of GP6, FYN, PKA, pPTK2, and pFAK1 in the CIS model group was markedly higher compared to the control group and anti-GP6 group. Protein kinases are enzymes that catalyze protein phosphorylation, which include an important protein kinase, protein kinase A (PKA).

**Limitations**

At present, a large number of studies have been carried out on the GP6 gene in China and abroad. The consistent view is that GP6 gene polymorphisms are related to platelet aggregation ability, but whether they lead to an increase or decrease in platelet aggregation, is still in question. If a polymorphism leads to an increase in platelet aggregation, then it is likely to be associated with thrombotic diseases. However, some scholars found that an increase in platelet aggregation, is still in question. If a polymorphism leads to an increase in platelet aggregation, then it is likely to be associated with thrombotic diseases. However, some scholars found that the polymorphism of GP6 is not significantly associated with the occurrence of coronary heart disease. Scholars generally agree that the expression of GP6 is enhanced in patients with acute ischemic diseases such as stroke and myocardial infarction. However, regarding whether GP6 gene polymorphisms are related to AS-based diseases (such as coronary heart disease and cerebral infarction), current research results are mostly negative and more in-depth studies are needed for verification. The safety of long-term drug use and the role of drug immunity and cardiovascular events need to be further explored. Chronic inflammation, which is repeated over a long time, can also lead to an inability to cure the disease. The presence of other pathways in the mechanism of GP6 in the treatment of AS requires further study.
Conclusions

**GP6** can promote the formation of cerebral ischemic stroke by upregulating the expression of **FCER1G**, **ITGA2**, **FYN**, **PKA**, **pPTK2**, and **pFAK1**. It may be a new treatment target for cerebral ischemic stroke.

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